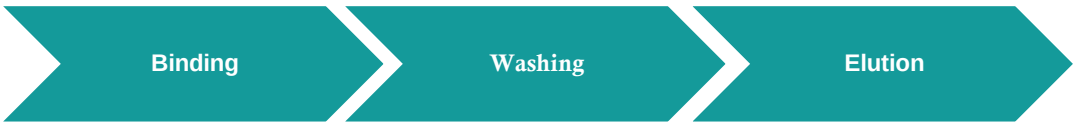
XpressPure PCR Clean up Kit

Protocol to recover or concentrate DNA fragments from PCR or other enzymatic reactions using XpressPure PCR Clean up Kit.

Process Flow



Kit Contents

Components	Storage Conditions	Shipping Conditions
XpressPure MagNa Mix	RT	RT
XpressPure Wash buffer	RT	RT
XpressPure Elution buffer	RT	RT

*RT denotes 15 - 25°C.

Materials not provided with the kit

1. 100% Ethanol to Wash Buffers as indicated on the bottle.

Important

Pay attention to standard lab practices and safety information before beginning the procedure. For more information, refer the appropriate Material Safety Data Sheet (MSDS) available from the product supplier or download from our website http://www.maggenome.com/

Technical Support

For any product related queries please write to us on info@maggenome.com, sales@maggenome.com, support@maggenome.com.

Sample Preparation

Single PCR amplicon or pooled PCR amplicons can be taken as the starting material. The protocol can be used for volumes from **10 µl - 100 µl.**

Protocol

DNA Binding	1. Vortex the **XpressPure MagNa Mix** to ensure complete dispersion of the particles. 2. Add **equal volume (1X) of XpressPure MagNa mix** to the starting material and gently pipette mix the contents for 10 -12 times. 3. Incubate the samples in RT for 15 minutes. 4. After incubation, place the tube on the MagNa Stand for 2 minutes or until the solution becomes clear. 5. Discard the supernatant without removing the tube from the MagNa Stand. Ensure the magnetic nanoparticles are not disturbed.

DNA Washing	6. Add **200 µl of XpressPure Wash buffer** and invert mix gently 5 - 6 times without removing the tube from the MagNa Stand and leave the wash buffer in the tube for at least 30 seconds. 7. Discard the supernatant without removing the tube from the MagNa Stand. 8. Repeat steps **6 - 7.** 9. Air dry the magnetic nanoparticles without removing the tube from MagNa Stand for at RT for 5 minutes. Avoid over drying.

DNA Elution	10. Remove the tube from the MagNa Stand and add **20 µl of XpressPure Elution Buffer.** 11. Carefully resuspend the magnetic nanoparticles by gentle pipette mixing. 12. Incubate the tube at RT for 10 minutes. 13. Place the tube on MagNa Stand for 5 minutes or until the solution becomes clear. 14. Carefully transfer the supernatant containing the purified amplicon to a fresh tube without removing the tube from MagNa Stand. Ensure the magnetic nanoparticles are not disturbed. 15. Discard the magnetic nanoparticles in appropriate hazard container.

Note: In the elution step, if MagNa particles take more than 10 min for clearing, then spin the tubes 14,000 rpm for 5 min, place on MagNa Stand and collect the supernatant for pure DNA.

Troubleshooting Guide

Observation	Possible causes	Suggested Solution
Low Yield	XpressPure MagNa Mix was improperly handled	Resuspend the MagNa Mix by vortexing prior to use. Pipette mix thoroughly while adding to samples.
	Magnetic nanoparticle loss during binding or washing steps	Carefully remove the supernatant from the tube without removing the tube from the MagNa Stand and without disturbing the Magnetic nanoparticles.
	Ethanol is not added to wash buffer	Add 100% ethanol to wash buffer before use as indicated on the bottle.
Poor performance in downstream sequencing applications	Ethanol carryover	Air dry the Magnetic nanoparticles after the washing steps to remove ethanol completely, but do not over dry the pellet.